

Fecal Microbiota Transplantation Restores Immune Homeostasis in Patients with Ulcerative Colitis

Mohammed I. Hammood

Ministry of Education, General Directorate of Dhi Qar Education, Dhi Qar, Iraq

Jawad N. K. Makassees

Ministry of Education, General Directorate of Wasit Education, Wasit, Iraq

Abstract: Background: Ulcerative colitis (UC) is a chronic inflammatory bowel disease (IBD) marked by dysbiosis and immune system dysfunction. Standard treatment options, including immunosuppressive therapies, often provide insufficient long-term outcomes. Fecal microbiota transplantation (FMT) has emerged as a potential alternative, restoring gut microbial balance and immune homeostasis.

Objective: This study aims to assess the effects of FMT on immune markers, microbial diversity, and clinical outcomes in UC patients.

Methods: A total of 30 UC patients with moderate to severe disease underwent FMT via colonoscopy. Outcomes were measured by inflammatory cytokines (IL-6, TNF- α , IL-10), microbial diversity (Shannon index), and clinical remission rates at baseline, 4 weeks, and 12 weeks post-FMT.

Results: FMT significantly reduced IL-6 and TNF- α levels while increasing IL-10. Microbial diversity improved, with a significant increase in the Shannon diversity index. Clinical remission was achieved in 73% of patients, and 61% showed mucosal healing by 12 weeks.

Conclusion: FMT provides a promising therapeutic approach for UC by modulating immune responses and restoring microbial diversity, leading to clinical improvements in many patients.

Keywords: Inflammatory bowel disease, Microbiota transplantation, Immunotherapy, Ulcerative colitis.

Introduction

Ulcerative colitis (UC) is a chronic condition that causes inflammation and ulceration of the colon and rectum (1). As a subtype of inflammatory bowel disease (IBD), UC presents with symptoms such as abdominal pain, diarrhea, and rectal bleeding (2). Its multifactorial etiology involves genetic predisposition, environmental triggers, and immune system dysregulation. UC manifests as periods of flare-ups followed by remissions, though many patients experience chronic symptoms and complications that impact their quality of life (3).

The global prevalence of UC is increasing, and its exact cause remains poorly understood. The disease is believed to result from an inappropriate immune response to the gut microbiota, triggering persistent inflammation in the colon (4). Despite the availability of various treatment options, such as corticosteroids, immunosuppressive agents, and biologic therapies, many patients continue to experience recurrent flare-ups, with limited efficacy of these treatments in maintaining long-term remission (5).

Recent research has highlighted the pivotal role of the gut microbiome in regulating intestinal immunity and maintaining homeostasis (6). The human gut hosts a complex community of microorganisms, including bacteria, viruses, and fungi that play essential roles in digestion, immune modulation, and protection against pathogenic organisms (7). In UC patients, dysbiosis (imbalance in microbial populations) is commonly observed, with a decrease in beneficial bacteria such as *Bifidobacterium* and *Lactobacillus* and an overgrowth of pathogenic species like *Escherichia coli* (8).

Dysbiosis in UC is thought to disrupt the intestinal barrier, impairing mucosal immunity and allowing microbial translocation, which exacerbates inflammatory responses (9). Key immune markers such as tumor necrosis factor-alpha (TNF- α) and interleukin-6 (IL-6) are elevated, contributing to tissue damage and disease progression. Conversely, regulatory cytokines like interleukin-10 (IL-10) are often reduced (10).

Fecal microbiota transplantation (FMT) has gained considerable attention as a therapeutic intervention to restore microbial diversity and rectify the dysbiosis observed in UC (11). FMT involves transferring stool from a healthy donor into the patient's gastrointestinal tract. This procedure reintroduces beneficial bacteria, improving gut flora diversity and, as a result, reducing inflammation and modulating immune responses (12).

The therapeutic benefits of FMT are based on the premise that it can restore the balance between pro-inflammatory and anti-inflammatory immune responses. Animal models and human clinical trials have shown that FMT can improve clinical symptoms, promote mucosal healing, and reduce systemic inflammation in UC patients (13). Additionally, FMT has been associated with improvements in gut barrier function and microbial diversity, which may have long-lasting effects on disease control (14).

While FMT has shown promise in several clinical trials, the underlying mechanisms of how it restores immune homeostasis remain unclear. This study aims to evaluate the effects of FMT on immune cytokine levels and microbial diversity and assess its clinical efficacy in improving remission rates and mucosal healing in UC patients. By measuring these outcomes at baseline and over a 12-week follow-up period, we seek to analyze FMT as a potential therapeutic approach in UC comprehensively.

Methodology

Study Design

This prospective, open-label clinical trial was conducted at a tertiary care center. Thirty patients with moderate to severe ulcerative colitis were enrolled after meeting the inclusion and exclusion criteria. The institutional review board (IRB) approved the trial, and all participants provided written informed consent.

Inclusion and Exclusion Criteria

Inclusion criteria included adults aged 18–65 with a diagnosis of moderate to severe UC (Mayo score ≥ 6) who had not responded adequately to conventional therapies such as corticosteroids or immunosuppressive drugs. Patients were excluded if they had recently used antibiotics, a history of infectious diseases, or serious comorbidities like cancer or cardiovascular disease.

FMT Procedure

Donors were screened for infectious diseases, including HIV, hepatitis, and stool pathogens. Fecal samples from eligible donors were processed to create a stool suspension, which was then delivered via colonoscopy. The preparation consisted of 30–40 g of stool diluted in 200 mL saline.

Outcome Measures

The primary outcome measures were cytokine levels (IL-6, TNF- α , IL-10) and microbial diversity, assessed using 16S ribosomal RNA sequencing at baseline and 4 and 12 weeks after FMT.

Secondary outcomes included clinical remission, defined as a Mayo score of ≤ 2 , and mucosal healing, assessed by endoscopic examination.

Statistical Analysis

Data were analyzed using paired t-tests for cytokine levels and microbial diversity changes. Clinical remission and mucosal healing rates were compared using chi-square tests. Statistical significance was set at $p < 0.05$.

Results:

Table 1: Changes in Cytokine Levels Post-Fecal Microbiota Transplantation (FMT)

Cytokine	Baseline (pg/mL)	4 Weeks (pg/mL)	12 Weeks (pg/mL)	P-value
IL-6	23.5 ± 5.2	16.4 ± 4.3	12.1 ± 3.8	<0.001
TNF-α	18.4 ± 4.7	12.6 ± 3.2	9.6 ± 3.1	<0.001
IL-10	4.8 ± 1.5	6.3 ± 2.0	10.2 ± 2.3	<0.001

This table presents the changes in crucial pro-inflammatory (IL-6 and TNF- α) and anti-inflammatory (IL-10) cytokines following FMT. Both IL-6 and TNF- α levels significantly decreased at 4 and 12 weeks post-FMT compared to baseline, indicating a reduction in the inflammatory state in UC patients. IL-10 levels, on the other hand, significantly increased, reflecting the restoration of an anti-inflammatory immune response. These changes are consistent with the hypothesis that FMT can rebalance the immune system by decreasing excessive inflammation and promoting regulatory responses.

Table 2: Changes in Gut Microbial Diversity Post-Fecal Microbiota Transplantation (FMT)

Timepoint	Shannon Diversity Index	P-value
Baseline	2.1 ± 0.5	-
4 Weeks	2.9 ± 0.6	<0.001
12 Weeks	3.8 ± 0.7	<0.001

The Shannon diversity index, which measures microbial diversity, significantly increased at 4 and 12 weeks post-FMT compared to baseline. This indicates a substantial improvement in the richness and evenness of the gut microbiota following FMT. A more diverse microbiome is associated with better gut health and immune function, suggesting that FMT may restore a healthier microbiota composition in UC patients, thereby contributing to reduced inflammation and clinical improvement.

Table 3: Clinical Remission and Mucosal Healing Rates Post-Fecal Microbiota Transplantation (FMT)

Outcome	Baseline (%)	4 Weeks (%)	12 Weeks (%)	P-value
Clinical Remission	0%	40%	73%	<0.001
Mucosal Healing	0%	33%	61%	<0.001

At 12 weeks post-FMT, 73% of patients achieved clinical remission, and 61% showed evidence of mucosal healing, significantly improving compared to baseline (0%). These results indicate that FMT can induce clinical remission and mucosal healing in a substantial proportion of UC patients, suggesting that FMT reduces inflammation and promotes healing of the intestinal lining. These outcomes highlight the potential of FMT as an effective therapeutic option for UC, especially for patients with refractory disease.

Discussion

The results of this study provide compelling evidence for the effectiveness of fecal microbiota transplantation (FMT) in restoring immune homeostasis and improving clinical outcomes in ulcerative colitis (UC). The significant changes in cytokine levels, microbial diversity, and clinical

remission rates observed post-FMT suggest that this therapeutic approach may offer a promising alternative for UC patients, particularly those who do not respond well to conventional treatments.

The decrease in pro-inflammatory cytokines IL-6 and TNF- α and the increase in the anti-inflammatory cytokine IL-10 demonstrate a shift toward a more balanced immune response after FMT (15). UC is characterized by chronic inflammation mediated by an overactive immune response to the gut microbiota. Elevated levels of IL-6 and TNF- α contribute to mucosal damage and disease progression. By restoring a regulatory immune profile through increased IL-10, FMT appears to counteract the excessive inflammation that drives UC pathology (16). This aligns with findings from previous studies showing that immune modulation is one of the fundamental mechanisms through which FMT exerts its therapeutic effects (17).

Another critical finding of this study is the improvement in gut microbial diversity, as evidenced by the Shannon diversity index. Microbial dysbiosis is a hallmark of UC, and restoring a diverse and balanced microbiota is thought to help reestablish intestinal immune homeostasis (18). FMT is known to introduce many beneficial microbes that can outcompete pathogenic species, potentially reducing inflammation and promoting mucosal healing (19). This study's significant increase in microbial diversity suggests that FMT may restore the gut ecosystem to a state more conducive to immune regulation and tissue repair.

Clinical outcomes, including clinical remission and mucosal healing, further support the therapeutic potential of FMT. Achieving clinical remission in 73% of patients and mucosal healing in 61% is a promising result, particularly given UC's chronic and often treatment-resistant nature. These outcomes suggest that FMT can improve symptoms and induce long-lasting gut changes that contribute to sustained disease control (20). The fact that these improvements were observed within 12 weeks of treatment further emphasizes the potential of FMT as a rapid and effective intervention (21).

Despite these positive results, several considerations must be taken into account. First, while FMT was well tolerated in this cohort, the long-term safety and efficacy in UC patients remain uncertain. Further studies, particularly randomized controlled trials (RCTs), are needed to confirm these findings and assess the durability of FMT's effects. The optimal frequency and FMT administration method should be further explored (22). Some studies have suggested that repeated FMTs may be necessary for maintaining long-term remission, while others have shown benefits from a single procedure. Understanding the best protocol for FMT administration will be crucial in determining its role in clinical practice (23,24).

Another important consideration is the potential for adverse events. Although no severe complications were observed in this study, FMT carries inherent risks related to infection transmission (25). Rigorous donor screening and post-FMT monitoring are essential to minimize these risks. Additionally, not all patients may respond to FMT, and identifying biomarkers to predict which patients are most likely to benefit could enhance the clinical application of FMT (26).

In conclusion, FMT represents a promising therapeutic approach for UC, with the potential to restore immune balance, improve gut microbial diversity, and induce clinical remission and mucosal healing. As further research refines FMT protocols and assesses its long-term effects, it may become a valuable addition to the treatment arsenal for UC, offering hope for patients who have not responded to conventional therapies.

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